

Rotational-Echo Double-Resonance NMR Distance Measurements for the Tubulin-Bound Paclitaxel Conformation

Younkee Paik,[‡] Chao Yang,[†] Belhu Metaferia,[†] Shoubin Tang,[†] Susan Bane,[‡] Rudravajhala Ravindra,[‡] Natasha Shanker,[‡] Ana A. Alcaraz,[§] Scott A. Johnson,[§] Jacob Schaefer,[‡] Robert D. O'Connor,[‡] Lynette Cegelski,[‡] James P. Snyder,[§] and David G. I. Kingston*[†]

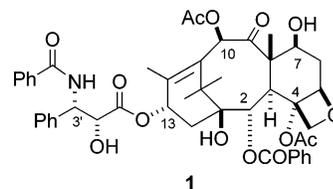
Contribution from the Department of Chemistry, M/C 0212, Virginia Polytechnic Institute and State University, Blacksburg, Virginia 24061, Department of Chemistry, State University of New York at Binghamton, Binghamton, New York 13902, Department of Chemistry, Emory University, Atlanta, Georgia 30322, and Department of Chemistry, Washington University, St. Louis, Missouri 63130

Received August 14, 2006; E-mail: dkingston@vt.edu

Abstract: The important anticancer drug Taxol (paclitaxel, PTX) owes its unique activity to its ability to bind to tubulin in a stoichiometric ratio and promote its assembly into microtubules. The conformation of the microtubule-bound drug has been the focus of numerous research efforts, since the inability of polymerized tubulin to form crystals precludes structure proof by X-ray crystallography. Likewise, although the α,β -tubulin dimer structure has been solved by electron crystallography, the 3.7 Å resolution is too low to permit direct determination of either ligand conformation or binding pose. In this article, we present experimental results from $^2\text{H}\{^{19}\text{F}\}$ REDOR NMR that provide direct confirmation that paclitaxel adopts a T-shaped conformation when it is bound to tubulin.

Introduction

The natural product paclitaxel (PTX) (Taxol, **1**) is a major cancer chemotherapeutic agent that is in clinical use for the treatment of breast, lung, and ovarian carcinomas.¹ Its importance is underscored by the fact that a number of analogues with improved activities have been prepared, some of which are in clinical trials.² PTX acts as a microtubule stabilizer, binding to microtubules and blocking microtubule dynamics, ultimately leading to damage to the mitotic spindle and apoptosis.³ Although PTX has other biological effects, its clinical activity is believed to be directly related to its microtubule-binding activity.^{3e,4}



One aspect of the binding of PTX to microtubules that is still being subjected to testing is the conformation of the various side chains in the bound molecule. Although PTX has a relatively rigid tetracyclic ring system, it possesses four flexible side chains at C2, C4, C10, and C13, and the presence of these freely rotatable units gives rise to numerous possible conformations. A detailed understanding of the microtubule-binding conformation of PTX could in principle lead to the development of improved and perhaps greatly simplified analogues. The elucidation of this conformation is thus a matter of potential practical as well as theoretical significance.

The direct approach to determine the binding conformation of the microtubule-PTX complex by X-ray crystallography is not available because of the polymeric and noncrystalline nature of the complex. Although it has proved possible to determine the structure of the tubulin dimer at 3.7 Å by electron crystallography of PTX-stabilized zinc-induced tubulin sheets,⁵ the structure lacks the resolution to define the detailed conformation of PTX on the tubulin polymer. It is thus necessary to use indirect methods to determine the bound conformation of

[†] Virginia Polytechnic Institute and State University.

[‡] State University of New York at Binghamton.

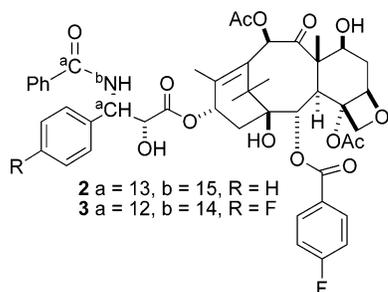
[§] Emory University.

[‡] Washington University.

- (1) (a) Rowinsky, E. K. *Annu. Rev. Med.* **1997**, *48*, 353–374. (b) Crown, J.; O'Leary, M. *Lancet* **2000**, *355*, 1176–1178. (c) Crown, J.; O'Leary, M.; Ooi, W.-S. *Oncologist* **2004**, *9* (Suppl. 2), 24–32. (d) Mekhail, T.; Markman, M. *Expert Opin. Pharmacother.* **2002**, *3*, 755–766.
 (2) (a) Kingston, D. G. I. In *Anticancer Agents from Natural Products*; Cragg, G. M., Kingston, D. G. I., Newman, D. J., Eds.; CRC Press: Boca Raton, FL, 2005; pp 89–122. (b) Geney, R.; Chen, J.; Ojima, I. *Med. Chem.* **2005**, *1*, 125–139. (c) Fang, W.-S.; Liang, X.-T. *Mini-Rev. Med. Chem.* **2005**, *5*, 1–12. (d) Zefirova, O. N.; Nurieva, E. V.; Ryzhov, A. N.; Zyk, N. V.; Zefirov, N. S. *Russ. J. Org. Chem.* **2005**, *41*, 315–351.
 (3) (a) Schiff, P. B.; Fant, J.; Horwitz, S. B. *Nature* **1979**, *277*, 665–667. (b) Wilson, L.; Jordan, M. A. *Nat. Rev. Cancer* **2004**, *4*, 253–265. (c) Horwitz, S. B. *Trends Pharmacol. Sci.* **1992**, *13*, 134–136. (d) Jordan, M. A.; Toso, R. J.; Thrower, D.; Wilson, L. *Proc. Natl. Acad. Sci. U.S.A.* **1993**, *90*, 9552–9556. (e) Blagosklonny, M. V.; Fojo, T. *Int. J. Cancer* **1999**, *83*, 151–156.
 (4) Blagosklonny, M. V.; Giannakakou, P.; El-Deiry, W. S.; Kingston, D. G. I.; Higgs, P. I.; Neckers, L.; Fojo, T. *Cancer Res.* **1997**, *57*, 130–135.

- (5) Lowe, J.; Li, H.; Downing, K. H.; Nogales, E. *J. Mol. Biol.* **2001**, *313*, 1045–1057.

PTX. Standard solution-state NMR methods cannot be used for microtubule-bound PTX due to the aqueous insolubility of the polymeric receptor. Nonetheless, recent advances in NMR spectroscopy carry the promise of exploiting spin diffusion to determine relative binding modes of microtubule-promoting ligands, including baccatin III, competing for the same binding site.⁶ In an alternative approach, solution-state NMR analysis of PTX conformations in the absence of the protein coupled with molecular dynamics simulations and the electron crystallographic (EC) density of the tubulin-PTX complex led to the proposal of T-Taxol as the bound conformation.⁷



Most previous studies of the bioactive conformation have used evidence from solution NMR analyses and molecular modeling and have led to the proposal of three different models for the solution conformation of PTX. NMR studies in nonpolar solvents suggested a “nonpolar” conformation with hydrophobic interactions between the C2 benzoate and the C3'-benzamido group.⁸ A “polar” or hydrophobic collapsed conformation was then proposed on the basis of NMR studies in polar solvents; this conformation features hydrophobic interactions between the C2 benzoate, the C3' phenyl group, and the C4 acetate.^{9–12} More recent studies using the NAMFIS deconvolution approach have shown that PTX adopts 9 to 10 conformations in CDCl₃,¹³ none of which achieves a population above 30%.

Although studies of the solution-phase conformations of PTX have been instructive, they cannot furnish direct information on the microtubule-bound conformation. The determination of the latter requires direct observation of microtubule-bound PTX, which necessitates the use of a technique that will permit measurement of internuclear distances on the ligand of a PTX-microtubule complex. Fortunately, rotational-echo double resonance (REDOR)¹⁴ allows just this measurement. One study using this technique has appeared to date. This study used the

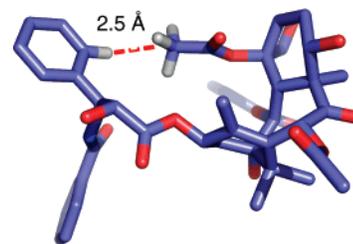


Figure 1. T-Taxol conformation illustrating the short C–H distance between the centroid of the C4 acetate group and the *ortho*-hydrogen of the C3' phenyl moiety.

2-(*p*-fluorobenzoyl)PTX **2** with ¹³C labels at the C3' amide carbonyl and C3' methine carbons and yielded F–¹³C distances of 9.8 and 10.3 Å between the fluorine and the C3' amide carbonyl and C3' methine carbons, respectively.^{15,16}

Analysis of the existing NMR data, both solution-phase and REDOR, in conjunction with knowledge of the EC density of β-tubulin led to the postulation of the T-Taxol conformation as the microtubule-binding conformation.^{7,17} Subsequently, a slightly different structure consistent with the REDOR data was proposed as the bioactive tubulin-binding conformation of PTX.¹⁸ However, the same structure is inconsistent with the EC density for PTX at the tubulin-taxane binding site.^{17,19} The T-Taxol conformation has been strongly supported by indirect evidence showing that the synthesis of constrained analogues based on this conformation yields compounds with improved activity.²⁰ In this article, we provide data on the synthesis and REDOR NMR analysis of additional PTX analogues which provide further experimental evidence for the T-Taxol conformation.

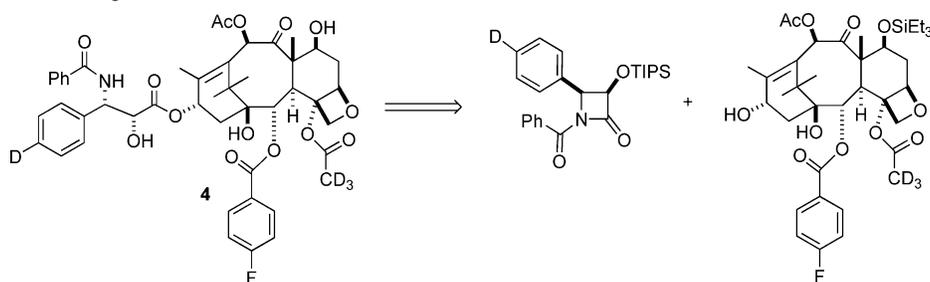
Results and Discussion

Design and Synthesis of PTX Analogues for REDOR Analysis. An analysis of the proposed bioactive T-Taxol conformation (Figure 1)^{7,17} indicated that the C3' phenyl group on the side chain was positioned quite close to the methyl group of the C4 acetate unit. This observation, in fact, was the inspiration for the synthesis of highly active bridged taxanes matching and, in some cases, surpassing the activity of parent PTX.²⁰ A critical test of the T-Taxol conformation would determine the distance between these two groups at the tubulin-taxane binding site. Unfortunately, the previous REDOR studies, with PTX analogues **2** and **3** labeled on the C2 benzoate and on the C13 side chain, provided no information on the spatial separation of the C4 acetate methyl group and atoms or groups located in other side chains.

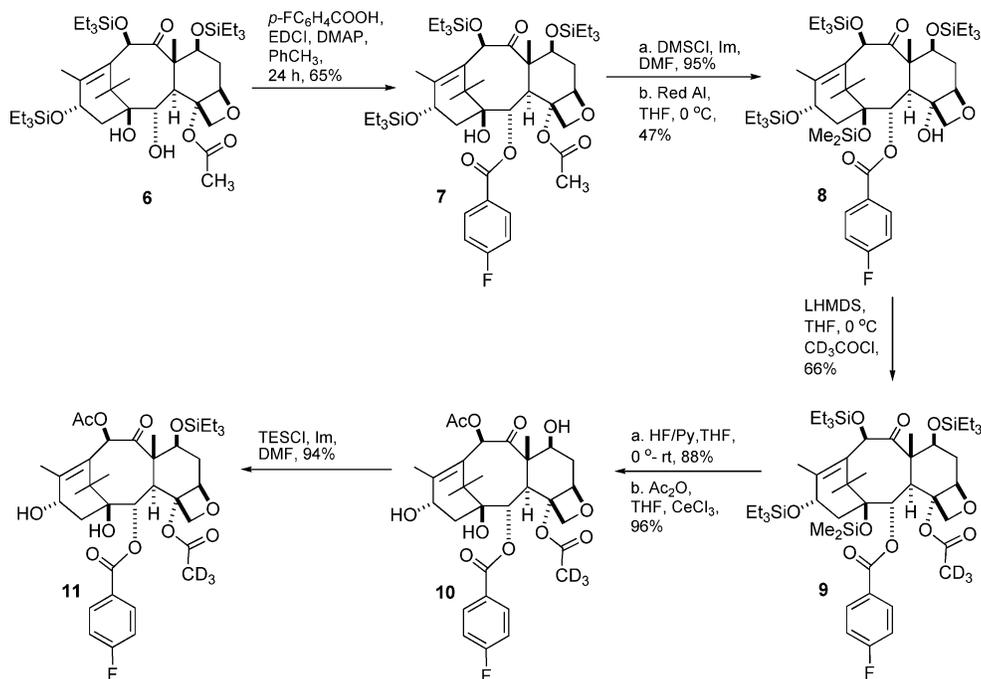
- (6) Sánchez-Pedregal, V. M.; Reese, M.; Meiler, J.; Blommers, M. J. J.; Griesinger, C.; Carlomagno, T. *Angew. Chem., Int. Ed.* **2005**, *44*, 4172–4175.
- (7) Snyder, J. P.; Nettles, J. H.; Cornett, B.; Downing, K. H.; Nogales, E. *Proc. Natl. Acad. Sci. U.S.A.* **2001**, *98*, 5312–5316.
- (8) (a) Dubois, J.; Guenard, D.; Gueritte-Voeglein, F.; Guedira, N.; Potier, P.; Gillet, B.; Betoil, J.-C. *Tetrahedron* **1993**, *49*, 6533–6544. (b) Williams, H. J.; Scott, A. I.; Dieden, R. A.; Swindell, C. S.; Chirlian, L. E.; Francl, M. M.; Heerding, J. M.; Krauss, N. E. *Can. J. Chem.* **1994**, *72*, 252–260. (c) Cachau, R. E.; Gussio, R.; Beutler, J. A.; Chmurny, G. N.; Hilton, B. D.; Muschik, G. M.; Erickson, J. W. *Supercomput. Appl. High Perform. Comput.* **1994**, *8*, 24–34.
- (9) Vander Velde, D. G.; Georg, G. I.; Grunewald, G. L.; Gunn, C. W.; Mitscher, L. A. *J. Am. Chem. Soc.* **1993**, *115*, 11650–11651.
- (10) Paloma, L. G.; Guy, R. K.; Wrasidlo, W.; Nicolaou, K. C. *Chem. Biol.* **1994**, *1*, 107–112.
- (11) Ojima, I.; Chakravarty, S.; Inoue, T.; Lin, S.; He, L.; Horwitz, S. B.; Kuduk, S. C.; Danishefsky, S. J. *Proc. Natl. Acad. Sci. U.S.A.* **1999**, *96*, 4256–4261.
- (12) Ojima, I.; Kuduk, S. D.; Chakravarty, S.; Ourevitch, M.; Begue, J.-P. *J. Am. Chem. Soc.* **1997**, *119*, 5519–5527.
- (13) Snyder, J. P.; Nevins, N.; Cicero, D. O.; Jansen, J. *J. Am. Chem. Soc.* **2000**, *122*, 724–725.
- (14) Gullion, T.; Schaefer, J. *Adv. Magn. Reson.* **1989**, *13*, 58–82.

- (15) Li, Y.; Poliks, B.; Cegelski, L.; Poliks, M.; Gryczynski, Z.; Piszczek, G.; Jagtap, P. G.; Studelska, D. R.; Kingston, D. G. I.; Schaefer, J.; Bane, S. *Biochemistry* **2000**, *39*, 281–291.
- (16) A related study, which has not yet been reported in full, used the radio frequency driven dipolar recoupling technique. This work indicated a distance of 6.5 Å between the fluorines of 2-(*p*-fluorobenzoyl)-3'-(*p*-fluorophenyl)-10-acetyldocetaxel (**3**), but it is not clear that this result is directly comparable to the REDOR results reported here: Ojima, I.; Inoue, T.; Chakravarty, S. *J. Fluorine Chem.* **1999**, *97*, 3–10.
- (17) Alcaraz, A. A.; Mehta, A. K.; Johnson, S. A.; Snyder, J. P. *J. Med. Chem.* **2006**, *49*, 2478–2488.
- (18) Geney, R.; Sun, L.; Pera, P.; Bernacki, R. J.; Xia, S.; Horwitz, S. B.; Simmerling, C. L.; Ojima, I. *Chem. Biol.* **2005**, *12*, 339–348.
- (19) Johnson, S. A.; Alcaraz, A.; Snyder, J. P. *Org. Lett.* **2005**, *7*, 5549–5552.
- (20) Ganesh, T.; Guza, R. C.; Bane, S.; Ravindra, R.; Shanker, N.; Lakdawala, A. S.; Snyder, J. P.; Kingston, D. G. I. *Proc. Natl. Acad. Sci. U.S.A.* **2004**, *101*, 10006–10011.

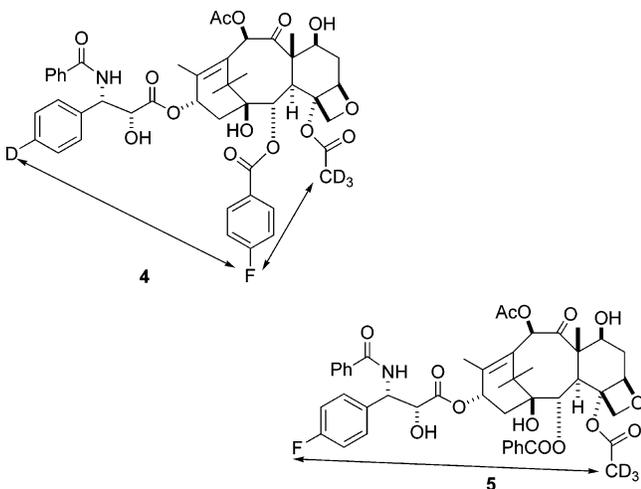
Scheme 1. Retrosynthesis of Ligand 4



Scheme 2. Synthesis of Labeled Baccatin 11



These considerations led to the design of the two PTX analogues 4 and 5 as ligands to determine key distances in



microtubule-bound PTX. Ligand 4 was particularly attractive, since in principle it allows the determination of two key distances; namely, between the C4 acetate and the C2 benzoate and between the C2 benzoate and the C3' phenyl. In the event, only a lower boundary on the latter distance could be determined for technical reasons, as discussed below. Ligand 5 was designed

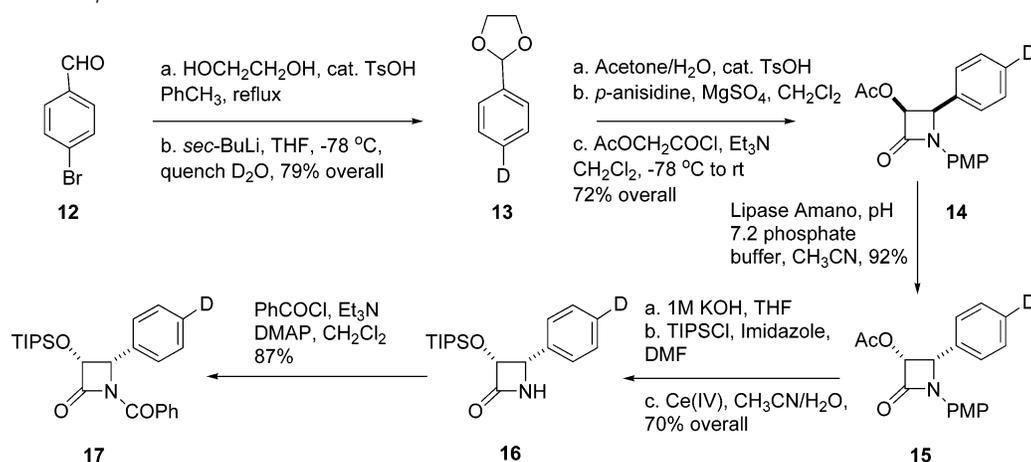
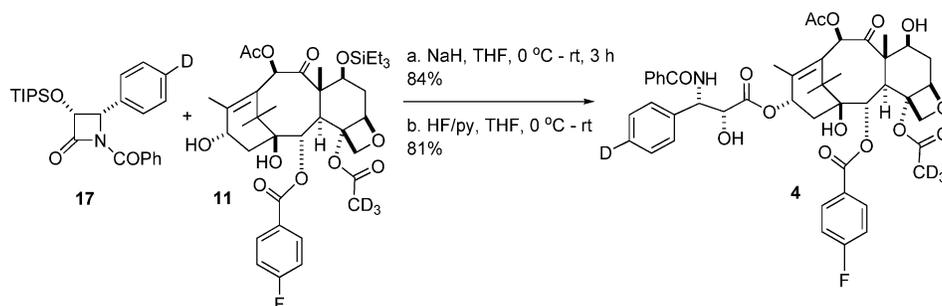
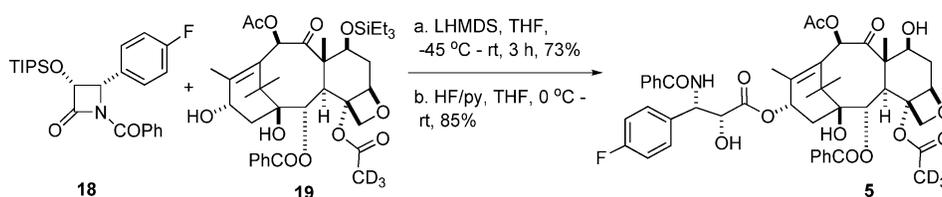
to allow a direct measurement of the distance between the C4 acetate and the C3' phenyl.

The labeled PTX were prepared from the available 10-deacetylbaccatin III. The retrosynthetic analysis of 4 is shown in Scheme 1.

The forward synthesis sequence is shown in Scheme 2. 10-Deacetylbaccatin III was converted to the C2 debenzoylated compound 6 by the literature procedure,²¹ and 6 was rebenzoylated to give 4-fluorobenzoate 7 in 65% yield. Deacetylation of the 4-acetate group by the published procedure²² gave the C4 alcohol 8, and this was reacylated with deuterated acetyl chloride to give the acetate 9 in 66% yield. The low yield from the acetylation reaction is attributed to partial loss of the dimethylsilyl protecting group and to incomplete conversion of the starting material, even after a prolonged reaction period (72 h). Global deprotection was followed by selective acetylation²³ of the 10-OH mediated by CeCl_3 to give compound 10. Selective reprotection of the 7-OH group completed the synthesis of the labeled baccatin core 11.

The syntheses of the required β -lactam derivative was achieved by literature procedures (Scheme 3).²⁴ The selected

- (21) Ojima, I.; Lin, S.; Inoue, T.; Miller, M. L.; Borella, C. P.; Geng, X.; Walsh, J. J. *J. Am. Chem. Soc.* **2000**, *122*, 5343–5353.
- (22) Chen, S.-H.; Kadow, J. F.; Farina, V.; Fairchild, C. R.; Johnston, K. A. *J. Org. Chem.* **1994**, *59*, 6156–6158.
- (23) Holton, R. A.; Zhang, Z.; Clarke, P. A.; Nadizadeh, H.; Procter, D. J. *Tetrahedron Lett.* **1998**, *39*, 2883–2886.

Scheme 3. Synthesis of β -Lactam **17****Scheme 4.** Synthesis of Labeled PTX Derivative **4****Scheme 5.** Synthesis of Labeled PTX Derivative **5**

starting material was 2-bromo benzaldehyde (**12**), which was converted to the deuterated acetal **13** and then to the deuterated β -lactam **14** through the *N*-(*p*-methoxyphenyl) (PMP) protected imines. Resolution of the (\pm)- β -lactam **14** was carried out with lipase PS (Amano) to yield the acetate of the desired (+)-enantiomer (**15**), along with the (–)-alcohols (not shown) in more than 92% combined yield. Functional group manipulations on **15** generated the triisopropylsilyl ether intermediate **16**, which was benzoylated to give the deuterated (+)-(3*R*,4*S*)-1-benzoyl-3-triisopropylsilyloxy-4-(*p*-deuterophenyl)azetid-2-one **17**.

The final steps of the synthesis of the target taxol analogue (**4**) consisted of a coupling reaction between the baccatin core (**11**) and β -lactam (**17**) followed by deprotection (Scheme 4).

Compound **5** was prepared by a similar route; the final steps are shown in Scheme 5, and full details of all synthetic procedures are included as Supporting Information.

Biological Evaluation of REDOR Analogues. The key activity criterion for the REDOR studies was the ability of the two compounds to promote tubulin assembly. In this respect, both analogues proved to be effective agents; compound **4** was about 2-fold less active than paclitaxel, while compound **5** was almost as active as paclitaxel (Table 1). Both compounds were

Table 1. Bioactivity of Paclitaxel and Analogues

compound	tubulin assembly activity, ED ₅₀ , μ M	A2780 cytotoxicity IC ₅₀ , μ M
1 (PTX)	1.7 \pm 0.4	0.023
4	3.5 \pm 0.7	7.6
5	2.6 \pm 0.5	0.047

also evaluated for cytotoxicity toward the A2780 ovarian cancer line. In this assay, the differences were more marked. The analogue **4** was more than 2 orders of magnitude less cytotoxic than paclitaxel against the A2780 ovarian cancer cell line, consistent with data on other similarly substituted analogues,²⁵ while analogue **5** was comparable in cytotoxicity to paclitaxel.²⁶

REDOR Spectra and ²H–¹⁹F Distances. A symmetrical, full quadrupolar spectrum (broken up into spinning sidebands)

(25) Kingston, D. G. I.; et al. *J. Med. Chem.* **1998**, *41*, 3715–3726.

(26) A reviewer asked whether there is any non-specific binding of the paclitaxel analogues to tubulin. We do not believe that there is any significant non-specific binding in these samples. The molar ratio of the ligand/tubulin dimer was 1:1, and the *K_d* for paclitaxel binding to microtubules is about 15 nM. Even if the affinities of the REDOR probes for the paclitaxel site on tubulin were 100-fold less than the affinity of paclitaxel for microtubules, > 95% of the probe would be bound to the protein under the conditions used to create the samples. Since the REDOR probes are nearly as active as paclitaxel as promoters of tubulin assembly, and since the structural perturbation between paclitaxel and the analogues is very small, it is reasonable to assume that the spectra are obtained on a homogeneous, fully bound sample.

(24) Brieva, R.; Crich, J. Z.; Sih, C. J. *J. Org. Chem.* **1993**, *58*, 1068–1075.

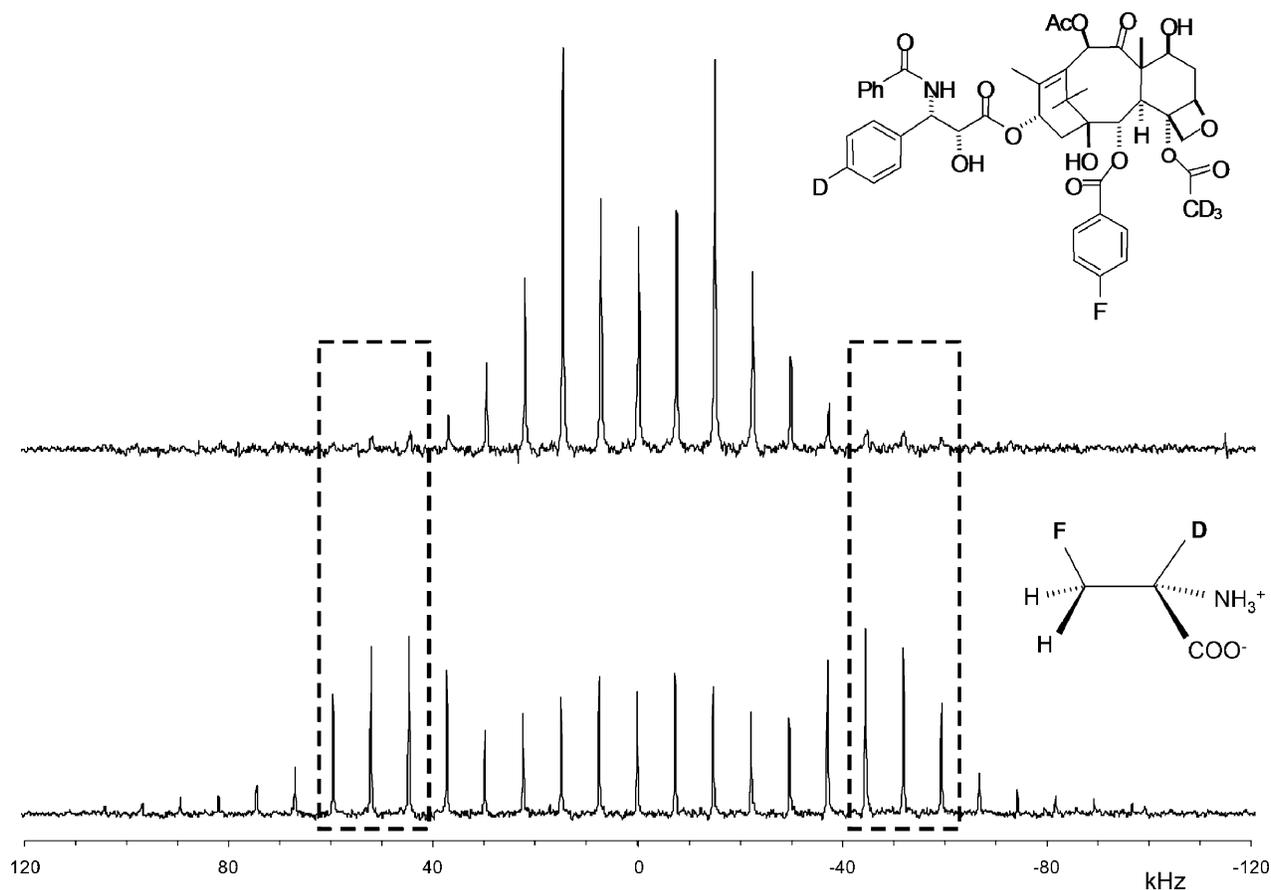


Figure 2. CPMAS ^2H NMR 2-T_1 Hahn-echo spectra of tubulin-bound paclitaxel analogue **4** (top) and D-[$2\text{-}^2\text{H},3\text{-}^{19}\text{F}$]alanine dispersed in a solid buffer (bottom). The aromatic ^2H spinning sidebands of the paclitaxel analogue are much weaker than the sidebands of the deuterated alanine (dotted boxes).

is observed for the D-[$2\text{-}^2\text{H},3\text{-}^{19}\text{F}$]alanine reference compound (Figure 2, bottom). This result indicates distortion-free ^2H detection over a bandwidth of more than 150 kHz. Although strong $n = \pm 8$ spinning sidebands are observed for the alanine reference, they are weak in the spectrum of tubulin-bound **4** (Figure 2, top); the strong central part of the spectrum of tubulin-bound **4** is due to its CD_3 group. We attribute this result to small-amplitude, low-frequency motions within the microtubule complex that interfere with echo formation for an aromatic ^2H with a large quadrupolar interaction. We believe that the motion is small amplitude because the cross-polarization magic-angle spinning (CPMAS) ^{19}F spectrum of tubulin-bound **5** is full strength, and the ^{19}F of **5** is on the same ring as the ^2H of **4**. Such motions are not present in the rigid lattice of the alanine crystal.

The 8% dephasing of the methyl- ^2H peak of tubulin-bound **4** after 64 rotor cycles (Figure 3) translates into an $\text{R}^1\text{--R}^2$ ($\text{CD}_3\text{--F}$) distance of $7.8 \pm 0.5 \text{ \AA}$ (Table 2). The $\text{R}^1\text{--R}^3$ distance ($\text{CD}_3\text{--F}$) in tubulin-bound **5** is $6.3 \pm 0.5 \text{ \AA}$ (Figure 4, inset, and Table 2).

The final key distance is the distance between R^2 and R^3 in tubulin-bound **4**. We expected an intensity ratio of methyl-to-aromatic ^2H peaks of 3:1 for tubulin-bound **4**, but instead observed a ratio of about 14:1 (Figure 3). This made quantitative determination of the dephasing of the aromatic ^2H peak difficult after 64 rotor cycles, and impractical for longer evolution times and even weaker signals. Nevertheless, we are confident that the 100% dephasing predicted by the polar model (Table 2) for an $\text{R}^2\text{--R}^3$ distance of 4.5 \AA is not present. The maximum

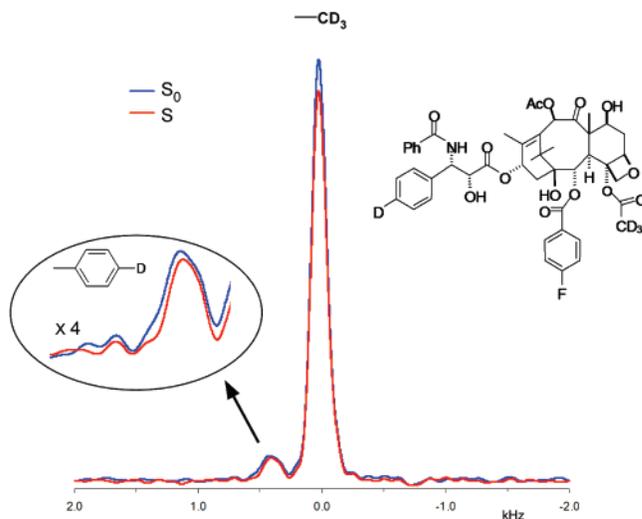


Figure 3. $^2\text{H}\{^{19}\text{F}\}$ REDOR spectra of tubulin-bound paclitaxel analogue **4** after 64 rotor cycles of dipolar evolution with magic-angle spinning at 8 kHz. The full-echo spectrum is shown in blue, and the dephased spectrum in red. The time domain signals were detected synchronously with the rotor so that all sidebands have been folded into the centerbands. The CD_3 peak is assigned zero frequency; the aromatic-D peak appears about 5 ppm to low field. The spectra resulted from the accumulation of 640 000 scans.

possible dephasing is less than 10% and is possibly much less than 10%. We conclude that the $\text{R}^2\text{--R}^3$ distance is greater than 8 \AA .

REDOR Quantitation. Our conclusions about $^2\text{H}\text{--}^{19}\text{F}$ distances from REDOR experiments depend on the efficiency

Table 2. Interatomic Distances for PTX Conformations As Compared to REDOR-Determined Separations for PTX on Tubulin

21

separation	compd	distances, Å					REDOR distance
		polar model ^a	nonpolar model ^b	PTX-NY model ^{c,d}	1JFF ^e	T-Taxol model ^{d,f}	
R ¹ –R ² ^g	4	7.4	8.0	7.3	6.5	7.9	7.8 ^h
R ¹ –R ³ ^g	5	5.5	7.2	6.4	7.2	6.6	6.3 ^h
R ² –R ³	4	4.5	12.5	13.1	11.6	12.2	>8 ^h
R ² –CH	2	9.6	8.5	9.4	9.3	9.9	10.3 ⁱ
R ² –C	2	10.4	6.2	10.0	8.1	9.1	9.8 ⁱ

^a Mastropaolo, D.; Camerman, A.; Luo, Y.; Brayer, G. D.; Camerman, N. *Proc. Natl. Acad. Sci. U.S.A.* **1995**, *92*, 6920–6924. ^b Williams, H. J.; Scott, A. I.; Dieden, R. A.; Swindell, C. S.; Chirlian, L. E.; Franci, M. M.; Heerding, J.; Krauss, N. E. *Tetrahedron* **1993**, *49*, 6545–6560; ref 13. ^c Reference 18. ^d The values of R²–CH and R²–C were inadvertently switched in Figure 1 of ref 17. ^e Reference 6. ^f Reference 7. ^g Distances from the 4-OAc methyl group were taken as an average of the three hydrogen positions. ^h This work, ± 0.5 Å. ⁱ Reference 15.

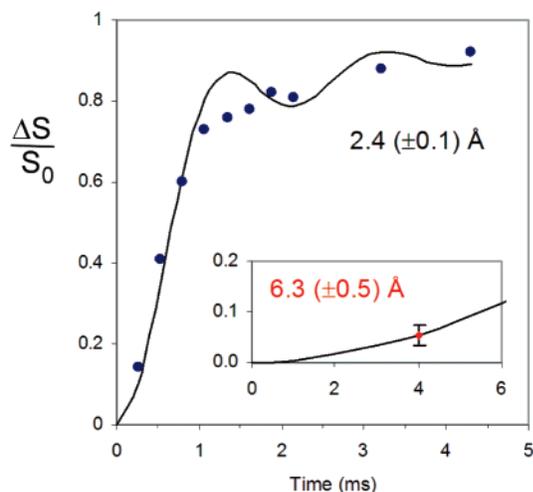


Figure 4. ${}^2\text{H}\{^{19}\text{F}\}$ REDOR dephasing ($\Delta S/S_0$) for the alanine sample of Figure 2. The experimental dephasing (●) is consistent with that calculated using SIMPSON (solid black line) assuming a single ${}^2\text{H}$ – ${}^{19}\text{F}$ distance of 2.4 Å. The corresponding REDOR spectra are shown in the Supporting Information (Figure S2). The inset shows the REDOR dephasing (red) for tubulin-bound paclitaxel analogue **5** after 32 rotor cycles of dipolar evolution with magic-angle spinning at 8 kHz. The REDOR spectra are shown in the Supporting Information (Figure S3). These spectra resulted from the accumulation of 1 056 000 scans. The experimental dephasing is consistent with that calculated using SIMPSON (solid black line) assuming a single ${}^2\text{H}_3$ – ${}^{19}\text{F}$ distance of 6.3 Å and the ${}^{19}\text{F}$ chemical shift and ${}^2\text{H}$ quadrupolar tensor parameters of Figure S4 of the Supporting Information. The minor effects of C_3 motional averaging were ignored.

of the REDOR dephasing pulses. We tested this efficiency using the alanine reference compound of Figure 2. The observed dephasing was close to 100% after 4 ms (Figure 4) and consistent with a three-bond distance in alanine.

The distance determinations also depend on the stability of the spectrometer under the long-term time averaging required to measure small differences between weak signals. For example, to determine the R¹–R³ distance in just 0.1 μmol of tubulin-bound **5** required more than 1 million scans and three weeks of spectrometer time. The stability of the spectrometer

for such an extended period was confirmed by performing an identical ${}^2\text{H}\{^{19}\text{F}\}$ experiment on a deuterated alanine sample that had no ${}^{19}\text{F}$ label. No REDOR difference was observed (Figure S3 of the Supporting Information). Short-term spectrometer stability in REDOR experiments is guaranteed by scan-to-scan alteration of the collection of dephased-echo (*S*) and full-echo (*S*₀) signals. Long-term stability arises from active control of the temperature-dependent drift in the power amplifiers.²⁷ This control eliminates fluctuations in pulse amplitudes that result in spectral artifacts. The drive voltages for the power amplifiers were determined by a control circuit that compares the amplitudes of calibration pulses that follow both *S* and *S*₀ acquisitions to a reference voltage. With this device, the power amplifiers were recalibrated once a second throughout the entire three-week data collection period.

The complete cancellation of *S* and *S*₀ illustrated in Figure S3 (bottom) was also aided by the properties of the transmission-line probe that was used for the measurements. The ${}^{19}\text{F}$ radio frequency (470.9 MHz) was introduced to the transmission line at an impedance minimum for the ${}^1\text{H}$ radio frequency (500.5 MHz). This provided about a 40 dB isolation between the tune-and-match circuits for the two frequencies²⁸ and a total isolation (including filters and traps) of almost 90 dB. There is no accidental leakage from one radio frequency channel to another in transmission-line probes of this design.

PTX Conformation Metrics; The Tubulin Bound Conformer. As mentioned earlier, three contenders for the bioactive bound form of PTX have received high level attention over the past decade: the nonpolar conformer, the polar form, and T-Taxol. The present solid-state REDOR-determined interatomic distances for labeled **4** and **5**, combined with complementary structural information, permit a distinction among these spatially distinct forms. The polar model proposed as the binding conformation in several reports^{8–10,15} is entirely consistent with the solid-state separations of R¹–R² and R¹–R³ recorded in

(27) Stueber, D.; Mehta, A. K.; Chen, Z.; Wooley, K. L.; Schaefer, J. *J. Polym. Sci., Part B: Polym. Phys.* **2006**, *44*, 2760–2775.

(28) Schaefer, J.; McKay, R. A. U.S. Patent 5,861,748, 1999.

Table 2. For all the PTX conformations discussed here, the constancy of R^1-R^2 is not surprising. The C4 acetate incorporating R^1 methyl can adopt three conformations. However, only that with the $C=O$ moiety positioned under the six-membered C-ring is observed in single-crystal X-ray structures. The same conformation is predicted by various computational methods to be lowest in energy. Similarly, while the C2–O bond of the C2 benzoyl group can theoretically adopt two conformations, only one is observed either experimentally (i.e., X-ray structures) or computationally. The second tucks the carbonyl group into the concave cavity of the baccatin core, causing severe steric compression. Consequently, adoption of the same conformations at C2 and C4 across the PTX conformational surface ensures a fixed R^1-R^2 as observed for the nonpolar, polar, and T-Taxol rotamers (Table 2). On the other hand, R^1-R^3 can be expected to vary since the C13 side chain can assume a variety of conformations.¹⁷ The polar and nonpolar models present distances that are -0.8 and $+0.9$ Å, respectively, from the REDOR value (Table 2). While these variations do not disqualify these conformers, they do highlight the fact that T-Taxol is within 0.3 Å of the R^1-R^3 measurement.

The R^2-R^3 phenyl–phenyl separation is crucial for rejecting the polar conformation as the bioactive form. The 4.5 Å gap falls far short of the >8 Å measure required by the REDOR experiment. By contrast, T-Taxol and the nonpolar conformers are compatible with this data point. In fact, both conformations would seem to match the trio of REDOR distances obtained in the present study reasonably well, an observation that might seem to confirm solution-phase NMR-based speculations that the nonpolar form is the bioactive one.⁷ The fundamental difference between them is the absence or presence, respectively, of hydrophobic collapse between the C2 and C3' *N*-benzoyl phenyl groups. As indicated by the structure associated with Table 2, the latter are unlabeled. As a consequence, potential REDOR dephasing experiments that might provide information on the association of the two C2 and C3' phenyl rings at the binding site (or the lack of it) are silent. On the other hand, our earlier REDOR study¹⁵ provides R^2-CH and R^2-C separations that offer a clear distinction. Since these distances have been discussed recently for other conformers,¹⁷ the focus here will be the nonpolar conformer. The latter structure's $F^2-^{13}CH$ and $F^2-^{13}C$ distances of 8.5 and 6.2 Å fall 1.8 and 3.6 Å, respectively, below the REDOR values (Table 2). The large discrepancy in distances is a consequence of the deep-seated repositioning of the C3' Ph and NHCOPh moieties from polar to nonpolar conformers. Furthermore, the results are in satisfying agreement with previously reported evidence suggesting that the C2 to C3' collapse is prevented by His227, which serves as the centerpiece of a sandwich incorporating the ligand phenyl groups on either side.^{7,20,29} A particularly compelling corroboration is the synthesis of bridged compounds holding the two phenyl groups in a collapsed orientation. Such compounds are inactive in the microtubule assembly assay and are unable to inhibit PTX binding.³⁰ Other more recent efforts to tether the

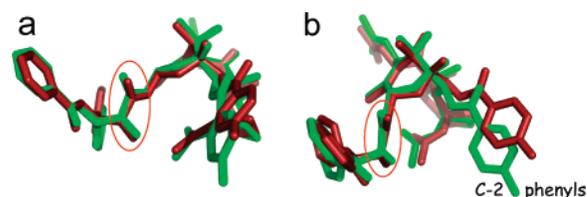


Figure 5. PTX-NY (green, ref 18) and T-Taxol (rust, ref 7) modified in four torsions of the C13 side chain as specified in Figure 5 of ref 18. Two different views (a and b). The key oxygens at C1' and C2' are circled.

PTX framework to a nonpolar architecture have likewise led to compounds with low or no activity.³¹ For these reasons, the nonpolar conformation would appear to be ruled out as a contender for the taxane bioactive conformation.

Two other conformations closely related to T-Taxol deserve mention. The first, the electron crystallographically refined structure 1JFF, is a close mimic of T-Taxol.^{7,17} In its conformational details, however, it is not quite as close a fit as the latter to the REDOR distances. Internal distances differ by 1.0 – 1.7 Å (Table 2). Thus, while the pdb-deposited low-resolution structure captures the essence of the bound conformation, the modeled refinement (T-Taxol)^{7,32} provides the best fit to the solid-state measurements. The second conformer, PTX-NY,¹⁸ is discussed in more detail below.

Analysis of the PTX-NY Conformation.³³ The PTX-NY conformation is a variation of the T-form that differs in the conformation of the C13 side chain from C1'–C3'. It matches all of the REDOR distances well (Table 2), and thus it is important to consider its candidacy for the paclitaxel bioactive conformation in the β -tubulin-taxane binding site. In previous articles, we ruled out PTX-NY as a candidate since our reconstruction of it strongly suggested that the conformer does not fit the EC density.^{17,19} Since the publication of these articles, Professor Ojima has kindly provided the original coordinates. This has made it possible to compare our reconstruction with the original and to perform two additional comparisons with it.

PTX-NY Reconstruction. Paclitaxel is a remarkably complex molecular skeleton with four-fused rings (A–D), 11 stereogenic centers, and four seemingly flexible side chains at C2, C4, C10, and C13. Nonetheless, the baccatin core (A–D rings) is rigid. The acetate at C10 is non-essential for bioactivity. C2 and C4 can, in principle, adopt a number of conformations, but, as described above, in practice each assumes only one in solution and in the solid state of several analogues. That leaves C13 as the primary flexible element in the molecule. However, apart from rotating phenyl rings, the conformation of this side chain is fully described by the following four dihedral angles: C12–C13–O13–C1', C13–O13–C1'–O1', O13–C1'–C2'–O2', and O2'–C2'–C3'–N3. For PTX-NY, the literature gives these angles as -160 , $+5$, $+20$, and $+175$ (ref 18, Figure 5). Accordingly, the corresponding four torsions in the T-Taxol conformation⁷ were changed to these values with no further alteration of the molecular structure. Figure 5 shows PTX-NY (green) overlaid on the four-angle modified T-Taxol (rust) by a five-point atom–atom superposition (i.e., C2, C11, C13, C15, and C3'). The terminal phenyl rings are offset somewhat, particularly at C2, but the conformation of PTX-NY is faithfully

(29) Kingston, D. G. I.; Bane, S.; Snyder, J. P. *Cell Cycle* **2005**, *4*, 279–289.

(30) Boge, T. C.; Wu, Z.-J.; Himes, R. H.; Vander Velde, D. G.; Georg, G. I. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 3047–3052.

(31) (a) Ojima, I.; Geng, X.; Miller, M. L.; Lin, S.; Pera, P.; Bernacki, R. J. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 349–352. (b) Geng, X.; Miller, M. L.; Lin, S.; Ojima, I. *Org. Lett.* **2003**, *5*, 3733–3736. (c) Querolle, O.; Dubois, J.; Thoret, S.; Dupont, C.; Guéritte, F.; Guénard, D. *Eur. J. Org. Chem.* **2003**, 542–550. (d) Querolle, O.; Dubois, J.; Thoret, S.; Roussi, F.; Montiel-Smith, S.; Guéritte, F.; Guénard, D. *J. Med. Chem.* **2003**, *46*, 3623–3630.

(32) Atomic coordinates of the computationally refined T-Taxol-tubulin structure can be found in the Supporting Information of ref 17.

(33) We thank a reviewer for suggesting that we add this section.

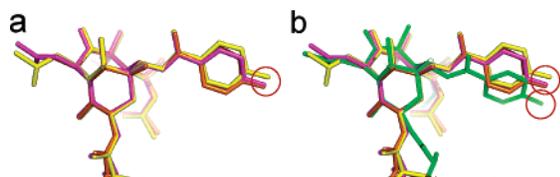


Figure 6. Taxane superpositions within the baccatin core. (a) Paclitaxel (magenta), docetaxel (orange), T-Taxol (yellow). (b) As in (a) with PTX-NY in green. The C13 side chain has been truncated at C2' to emphasize differences in the remainder of the molecule.

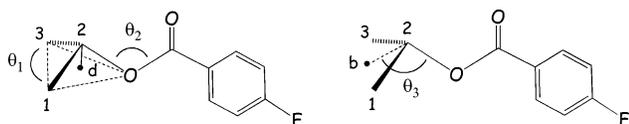


Figure 7. Geometry around C2 in the paclitaxel baccatin core: bond angles θ_1 (C1–C2–C3), θ_2 (C2–O–C(=O)), and θ_3 . The latter measures from the bisector (b) of C1 and C3 to the ester O (i.e., (b–C2–O)). Variable d corresponds to the distance from C2 to the centroid of C1, C3, and O.

Table 3. Geometry around C2 for Selected Taxanes As Defined in Figure 7; Degrees and Å

	θ_1	θ_2	θ_3	d
PTX X-ray	117.4	119.6	123.5	0.49
DTX X-ray	117.3	119.6	121.7	0.50
T-Taxol	119.1	119.1	123.9	0.48
PTX-NY	125.3	123.1	130.6	0.41

reproduced from C1' to C3' as highlighted by the circled atoms in Figure 5. As a result, the previous comparisons of conformations^{17,19} would appear to be valid.

C2 *para*-Fluorophenyl Orientation. All distances in Table 2 that include R² (four of five) involve measurement from a fluorine atom at the *para* position of the C2 benzoyl phenyl to other atoms in the taxane structure. Accurate atomic separations in a given structure require that the fluorine is correctly located relative to other atoms in the molecule. Given the spatial discrepancy of the C2 phenyl rings in Figure 5, we decided to examine which of the two structures is responsible and to try to understand the origin of the difference. As standards, we selected the X-ray structures of two landmark taxanes: Taxol³⁴ (PTX, paclitaxel, magenta) and taxotere³⁵ (DTX, docetaxel, orange). These were supplemented with *para*-F atoms and superposed by aligning the baccatin cores (i.e., atoms C2, C11, C13, and C15). T-Taxol (yellow) was treated similarly. Figure 6a shows that, in all three cases, the C2 ester group and the pendent phenyl rings overlap well; the three fluorines cluster within the red sphere with a radius of 1.1 Å.

The identical superposition of PTX-NY is shown in Figure 6b. It would appear that, while the remainder of the paclitaxel molecule is reasonably optimized, the C2 side chain in PTX-NY is at odds with the experimental structures. The origin of the displacement is captured in Figure 7 and quantified in Table 3.

While the T-Taxol geometry around C2 is very similar to the experimental structures, PTX-NY exhibits a distortion characterized by a flattening of the C2 tetrahedral geometry and by an expansion of θ_3 , the C2–O–C(=O) bond angle, to 131°.

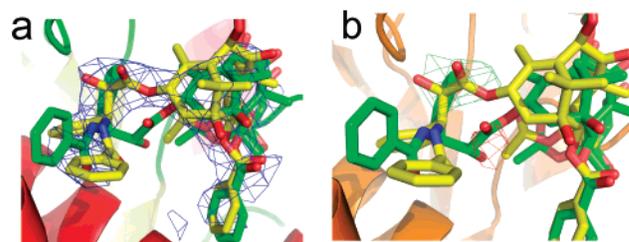


Figure 8. Structures of T-Taxol (yellow) and PTX-NY (green) superposed in the 1JFF tubulin-binding site. (a) The $2F_{\text{obsd}} - F_{\text{calcd}}$ omit map is shown as a blue 3D contour. (b) Difference maps ($F_{\text{obsd}} - F_{\text{calcd}}$) for the PTX-NY structure. Green corresponds to unfilled density, red to incorrectly filled density.

Both work together to displace the C2 side chain toward the concave face of the baccatin core, as illustrated by Figures 5b and 6b. The implication is that, while the PTX-NY structure closely matches the REDOR distances of Table 2, it is possible that the match for four of the five distances dependent on the location of the C2 aromatic-F is fortuitous and based on a geometry that deviates significantly from experiment.

Electron Crystallographic Density Fitting. Our attempts to fit PTX-NY into the electron crystallographically refined taxane-binding site^{5,7} without steric congestion have not been completely satisfying. Geney and co-workers apparently faced the same problem, since they report extensive rearrangement of the protein following docking of PTX-NY onto the DTX ligand-placeholder in unrefined 1TUB³⁶ and subsequent protein–ligand minimization. A risk in the latter procedure is that introducing a predetermined conformation into the binding cleft followed by minimization to relieve short nonbonded contacts can cause the system to deviate significantly from the experimental structure. However, to make the fairest possible comparison between PTX-NY and T-Taxol, we decided to employ the experimental structure (1JFF) in which both ligands are manually docked and superimposed (Figure 8), as previously shown by both Geney et al.¹⁸ and our group.^{17,19} In this way, the primary difference between them is emphasized; namely, the path taken by the C1' to C3' segment of the C13 side chain. Figure 8a illustrates that the trace of the EC density clearly encompasses the C1'–C3' atoms in T-Taxol, while the same atoms in PTX-NY are not centered in ligand density. Difference density maps for PTX-NY (Figure 8b) express this result another way. The green difference contour illustrates density unfilled by the ligand, while the red contour represents density that is incorrectly filled. By contrast, the T-Taxol conformer is in complete agreement with the same maps and in accord with very similar visualizations resulting from analysis of reconstructed PTX-NY (Figure 5).^{17,19}

Conclusions and Prospects

Paclitaxel is one of the unique blockbuster anticancer drugs to emerge from natural product screening in the past two decades. Analogues continue to be created and evaluated in the clinic for improved efficacy, reduced toxicity, and the ability to elude resistance. Firm knowledge of the conformation of the molecule bound to its tubulin–microtubule target would answer long-standing questions, while contributing to future efforts to remodel taxanes with the baccatin core or to completely re-engineer the molecule's architecture. Accordingly, we have

(34) Mastropaolo, D.; Camerman, A.; Luo, Y.; Brayer, G. D.; Camerman, N. *Proc. Natl. Acad. Sci. U.S.A.* **1995**, *92*, 6920–6924.

(35) Guéritte-Voegelein, F.; Guénard, D.; Mangatal, L.; Potier, P.; Guilhem, J.; Césario, M.; Pascard, C. *Acta Crystallogr., Sect. C* **1990**, *46*, 781–784.

(36) Nogales, E.; Wolf, S. G.; Downing, K. H. *Nature* **1998**, *391*, 199–203.

prepared a series of fluoro- and deuterio-labeled derivatives of PTX suitable for measurement of interatomic distances by solid-state $^2\text{H}\{^{19}\text{F}\}$ REDOR NMR. Samples of ligand-doped microtubules were subjected to three weeks of spectrometer time to measure each of three pairwise distances. The latter were compared to the same quantities for five PTX conformers, each of which has been proposed to be the bioactive form at the tubulin-binding pocket.

The outcome definitively eliminates the polar conformer from consideration. The nonpolar conformer is also eliminated by the combination of the present data with our previous REDOR data.¹⁵ The PTX-NY structure fits the REDOR data well, though there are irregularities in its molecular geometry, as illustrated by Figures 6 and 7 and Table 3. The latter alone, however, do not disqualify the conformer from bioactive candidacy. Critical is the observation that the shape of PTX-NY's C13 side chain is in conflict with the experimental EC density. On the other hand, the T-Taxol structure passes three fundamentally important tests. First, this conformation is within 0.7 Å of four well-defined REDOR distances and fully consistent with the fifth. Second, it is the only conformer in Table 2 that is compatible with the EC density. Third, it is unique in being a template for synthesis of bridged taxanes that match or surpass both the binding affinity and cytotoxicity characteristics of the paclitaxel parent. For these reasons, the T-Taxol conformation proposed by a combination of structure and modeling techniques and by synthesis and bioassay²⁰ is confirmed by REDOR measurement of interatomic distances.

Finally, with respect to evaluating prospective bioactive taxane conformations at the binding site of β -tubulin, it must be noted that the EC binding model that led to the T-Taxol proposal was derived from Zn^{+2} -stabilized tubulin sheets^{5,7} and not genuine microtubules. It is thus necessary to determine whether this model is suitable as a guide to the 3D-shape of the bioactive form at the tubulin-binding site. It is no coincidence that virtually every recent synthetic program attempting to prepare more potent and less resistant taxanes based on biostructural principles has employed the EC structure.^{18,20,29,37} Furthermore, the tubulin dimer structure derived from Zn^{+2} -stabilized sheets has been modeled into the 8 Å resolution microtubule structure determined by cryoelectron microscopy³⁸ and shown to be entirely consistent with the MT structure. Finally, as we have pointed out previously,^{17,20,29} not only is the REDOR geometry derived from lyophilized MTs co-incident with the T-Taxol form (Table 2), but also bioactivities in situations where MTs are present as soluble dynamic entities can be significantly improved by exploiting the EC structure and bound ligand from stabilized tubulin sheets. The implication is clear. While the details of tubulin polymerization may vary

on the level of protein assembly, the taxane-binding site sequestered within β -tubulin appears to be faithfully retained in the two situations. This underscores the necessity that alternative bioactive conformations for paclitaxel pass the three tests outlined in the previous paragraph.

Experimental Section

Synthesis of Labeled Paclitaxels. Full experimental details of the syntheses of compounds **4** and **5** are provided as Supporting Information.

ED₅₀ Determination. Pure tubulin was obtained from bovine brain and used in polymerization and REDOR experiments. Polymerization activity of paclitaxel and derivatives was evaluated in the following manner. GTP-tubulin (5 μM) in a buffer containing 0.1 M PIPES, 1 mM MgSO_4 , 2 mM EDTA, and 0.1 mM GTP (pH 6.90) was equilibrated to 37 °C in a quartz cuvette in a Hewlett-Packard model 8453 diode array spectrophotometer, and a baseline was recorded. Polymerization was initiated by addition of the compound to be tested in DMSO, and the extent of assembly was measured as the change in apparent absorption at 350 nm. In all the solutions, the percentage of DMSO was maintained as 4% v/v to avoid any effect of DMSO on tubulin assembly. A plot of change in apparent absorption at 350 nm versus the concentration of the compound was fit to a rectangular hyperbola to obtain the ED₅₀ value.

NMR Samples. Paclitaxel analogues **4** (0.7 μmoles) and **5** (0.1 μmoles) were bound 1:1 to tubulin to form microtubule complexes following the protocol described earlier¹⁴ with the following modifications: GMPCPP rather than GTP was employed as the nucleotide, and 20 mM sucrose and 1% PEG (M_n 8000) were used as the lyo- and cryoprotectants, respectively. A small amount of the freeze-dried material was reconstituted in 10 mM phosphate buffer (pH 7.0) and examined by electron microscopy; normal microtubules were observed. The complexes were packed into rotors under a dry atmosphere. A reference sample for testing $^2\text{H}\{^{19}\text{F}\}$ REDOR dephasing was prepared by dissolving D-[2- ^2H ,3- ^{19}F]alanine (10 mg) with HEPES buffer (100 mg), flash freezing the solution, and then lyophilizing at low temperature. A control sample that contained no ^{19}F was prepared by physically mixing 5 mg of D-[3- $^2\text{H}_3$]alanine (5 mg) with sulfur powder (100 mg). This mixture was used to confirm that a null REDOR difference signal was observed (Figure S3 of the Supporting Information) from a 32-T, $^2\text{H}\{^{19}\text{F}\}$ REDOR experiment after 1 million scans.

Spectrometer. Deuterium NMR spectra were obtained using a spectrometer equipped with a four-frequency (HFCD) transmission-line probe operating at an ^1H frequency of 500.5 MHz and an ^2H frequency of 76.83 MHz. Tune and match circuits for each frequency were external to the magnet. The probe had a 12-mm long, 6-mm inside-diameter analytical coil and a Chemagnetics/Varian magic-angle spinning ceramic stator. Five-millimeter thin-wall ceramic rotors were spun at 8000 Hz with the speed under active control to within ± 2 Hz. A 12-T static magnetic field was provided by an 89-mm bore MagneX superconducting solenoid. The spectrometer was controlled by a Tecmag Apollo pulse programmer. Levels of all radio frequency pulses were under active control during data acquisition by sampling of test pulses fired on each channel in use immediately after each scan.

REDOR. The version of REDOR used in these experiments (Figure S1) had a single ^2H 180° pulse (5.6 μs) and $16n$ ($n = 2, 3, \dots$) ^{19}F 180° pulses (5.8 μs). The phases of the ^{19}F pulses followed the xy8 phase-alternation scheme.³⁹ This combination gave the best echo refocusing and most complete dephasing (Figure S2). Internuclear distances were calculated from the REDOR dephasing by spectral simulations performed using SIMPSON.⁴⁰ The $^1\text{H} \rightarrow ^2\text{H}$ matched cross-polarization

- (37) (a) Boge, T. C.; Wu, Z.-J.; Himes, R. H.; Vander Velde, D. G.; Georg, G. I. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 3047–3052. (b) He, L.; Jagtap, P. G.; Kingston, D. G. I.; Shen, H.-J.; Orr, G. A.; Horwitz, S. B. *Biochemistry* **2000**, *39*, 3972–3978. (c) Giannakakou, P.; Gussio, R.; Nogales, E.; Downing, K. H.; Zaharevitz, D.; Bollback, B.; Poy, G.; Sackett, D.; Nicolaou, K. C.; Fojo, T. *Proc. Natl. Acad. Sci. U.S.A.* **2000**, *97*, 2904–2909. (d) Barboni, L.; Lambertucci, C.; Appendino, G.; Vander Velde, D. G.; Himes, R. H.; Bombardelli, E.; Wang, M.; Snyder, J. P. *J. Med. Chem.* **2001**, *44*, 1576–1587. (e) Querolle, O.; Dubois, J.; Thoret, S.; Roussi, F.; Guéritte, F.; Guénard, D. *J. Med. Chem.* **2004**, *47*, 5937–5944. (f) Hari, M.; Loganzo, F.; Annable, T.; Tan, X.; Musto, S.; Morilla, D. B.; Nettles, J. H.; Snyder, J. P.; Greenberger, L. M. *Mol. Cancer Ther.* **2006**, *5*, 270–278.
- (38) (a) Nogales, E.; Whittaker, M.; Milligan, R. A.; Downing, K. H. *Cell* **1999**, *96*, 79–88. (b) Li, H.; DeRosier, D. J.; Nicholson, W. V.; Nogales, E.; Downing, K. H. *Structure* **2002**, *10*, 1317–1328.

- (39) Gullion, T.; Baker, D. B.; Conradi, M. S. *J. Magn. Reson.* **1990**, *89*, 479–484.

- (40) Bak, M.; Rasmussen, J. T.; Nielsen, N. C. *J. Magn. Reson.* **2000**, *147*, 296–330.

transfer was at 54 kHz for 3 ms, with an ^1H 90° pulse of $2.5 \mu\text{s}$ and 100 kHz dipolar decoupling with TPPM⁴¹ phase modulation.

Acknowledgment. The work at Virginia Polytechnic Institute and State University, SUNY Binghamton, and Emory University was supported by the NIH Grant CA-69571, and the work at Washington University was supported by NIH Grant EB-001964; we are grateful for this support. We thank William Bebout (VPISU) for mass spectroscopic determinations and Andrew Norris and Peggy Brodie (VPISU) for A2780 cytotoxicity assays. We are likewise grateful to Professor Iwao Ojima for providing the CVFF coordinates to PTX-NY.

(41) Bennett, A. E.; Rienstra, C. M.; Auger, M.; Lakshmi, K. V.; Griffin, R. G. *J. Chem. Phys.* **1995**, *103*, 6951–6958.

Supporting Information Available: Full experimental details of the syntheses of labeled PTX analogues **4** and **5** and **S23**. Figures showing pulse sequence for $^2\text{H}\{^{19}\text{F}\}$ REDOR; $^2\text{H}\{^{19}\text{F}\}$ REDOR spectra of D-[2- ^2H ,3- ^{19}F]alanine dispersed in a solid buffer as a function of the dipolar evolution time; $^2\text{H}\{^{19}\text{F}\}$ REDOR spectra of tubulin-bound paclitaxel analogue **5** and $^2\text{H}\{^{19}\text{F}\}$ REDOR spectra of a control sample of D-[3- $^2\text{H}_3$]alanine; CPMAS ^{19}F and ^2H NMR spectra of tubulin-bound paclitaxel analogue **4**, and simulated spectra using SIMPSON. Complete ref 25. This material is available free of charge via the Internet at <http://pubs.acs.org>.

JA0656604